

Identification of 4 autophagy-related genetic variants as risk factors for chronic lymphocytic leukemia

Antonio José Cabrera-Serrano,^{1,2} José Manuel Sánchez-Maldonado,¹⁻³ Juan José Rodríguez-Sevilla,⁴ Fernando Jesús Reyes-Zurita,³ Rosa Collado,⁵ Anna Puiggros,^{6,7} Elena Cornejo-Calvo,^{2,8} Paloma García-Martín,^{2,8} Rob Ter Horst,^{9,10} Yolanda Benavente,^{11,12} Andrés Jerez,¹³ Stefano Landi,¹⁴ Blanca Espinet,^{6,7} Rossana Maffei,¹⁵ Miguel Ángel López-Nevo,^{2,16} Silvia Ramos-Campoy,^{6,7} Carmen González-Olmedo,¹⁷ Tzu-Hua Chen-Liang,¹⁸ Víctor Moreno,^{12,19,20} Fatin Jannus,³ Rafael Marcos-Gragera,^{12,21,22} María Carretero-Fernández,^{1,2} Belém Sampaio-Marques,²³ Irene Gámez,¹⁸ María García-Álvarez,²⁴ Nicola J. Camp,²⁵ Trinidad Dierssen-Sotos,^{12,26} Joanna Kamaso,^{6,7} Eva María Pérez,^{2,8} Aaron D. Norman,²⁷ Mario Luppi,¹⁵ Yang Li,^{9,28} Miguel Alcoceba,²² Daniele Campa,¹⁴ Silvia de San José,^{11,12} Roberto Marasca,¹⁵ Paula Ludovico,²³ Alyssa Clay-Gilmour,²⁹ Federico Canzian,³⁰ Marian Ibañez,⁵ Mihai G. Netea,^{9,31} James McKay,³² Delphine Casabonne,^{11,12} Sonja I. Berndt,³³ Susan L. Slager,³⁴ and Juan Sainz^{1-3,12}

¹Genomic Oncology Area, Centre for Genomics and Oncological Research, Pfizer/University of Granada/Andalusian Regional Government, Parque Tecnológico de la Salud, Granada, Spain; ²Instituto de Investigación Biosanitaria IBs.Granada, Granada, Spain; ³Department of Biochemistry and Molecular Biology I, Faculty of Sciences, University of Granada, Granada, Spain; ⁴Hematology Department, Hospital del Mar, Barcelona, Spain; ⁵Hematology Department, Hospital General de Valencia, Valencia, Spain; ⁶Molecular Cytogenetics Laboratory, Pathology Department, Hospital del Mar, Barcelona, Spain; ⁷Translational Research on Hematological Neoplasms Group, Cancer Research Program, Hospital del Mar Research Institute, Barcelona, Spain; ⁸Hospital Campus de la Salud, Parque Tecnológico de la Salud Granada, Granada, Spain; ⁹Department of Internal Medicine and Radboud Center for Infectious Diseases, Radboud University Medical Center, Nijmegen, The Netherlands; ¹⁰CeMM Research Center for Molecular Medicine of the Austrian Academy of Sciences, Vienna, Austria; ¹¹Catalan Institute of Oncology, Bellvitge Biomedical Research Institute, Consortium for Biomedical Research in Epidemiology and Public Health, University of Barcelona, Barcelona, Spain; ¹²CIBER Epidemiología y Salud Pública, Madrid, Spain; ¹³Experimental Hematology Unit, Department of Hematology, Vall d'Hebron Institute of Oncology, University Hospital, Vall d'Hebron, Barcelona, Spain; ¹⁴Department of Biology, University of Pisa, Pisa, Italy; ¹⁵Department of Medical and Surgical Sciences, University of Modena and Reggio Emilia, AOU Policlinico, Modena, Italy; ¹⁶Immunology Department, Virgen de las Nieves University Hospital, Granada, Spain; ¹⁷Medical Oncology Unit, University Hospital of Jaén, Jaén, Spain; ¹⁸Hematology Department, Morales Meseguer University Hospital, Murcia, Spain; ¹⁹Unit of Biomarkers and Susceptibility, Cancer Prevention and Control Program, Bellvitge Biomedical Research Institute, Catalan Institute of Oncology; ²⁰Department of Clinical Sciences, Faculty of Medicine, University of Barcelona, Barcelona, Spain; ²¹Epidemiology Unit and Girona Cancer Registry, Oncology Coordination Plan, Department of Health, Autonomous Government of Catalonia, Catalan Institute of Oncology, Girona Biomedical Research Institute (IdiBGi-CERCA), and Universitat de Girona, Girona, Spain; ²²Josep Carreras Leukemia Research Institute, Girona, Spain; ²³Life and Health Sciences Research Institute, School of Medicine, University of Minho, Braga, Portugal; ²⁴Department of Hematology, University Hospital of Salamanca (HUS/IBSAL), CIBERONC, Cancer Research Institute of Salamanca-IBMCC (USAL-CSIC), Salamanca, Spain; ²⁵Division of Hematology and Huntsman Cancer Institute, The University of Utah, Salt Lake City, UT; ²⁶University of Cantabria IDIVAL, Santander, Spain; ²⁷Department of Quantitative Health Sciences, Mayo Clinic, Rochester, MN; ²⁸Centre for Individualised Infection Medicine and TWINCORE, joint ventures between the Helmholtz-Centre for Infection Research and the Hannover Medical School, Hannover, Germany; ²⁹Department of Epidemiology and Biostatistics, Arnold School of Public Health, University of South Carolina, Greenville, SC; ³⁰Genomic Epidemiology Group, German Cancer Research Center, Heidelberg, Germany; ³¹Department for Immunology and Metabolism, Life and Medical Sciences Institute, University of Bonn, Bonn, Germany; ³²International Agency for Research on Cancer, Lyon, France; ³³Division of Cancer Epidemiology and Genetics, National Cancer Institute, Bethesda, MD; and ³⁴Department of Health Sciences Research, Mayo Clinic, Rochester, MN

Submitted 4 June 2025; accepted 29 July 2025; prepublished online on *Blood Advances* First Edition 3 September 2025; final version published online 26 November 2025. <https://doi.org/10.1182/bloodadvances.2025017345>.

Genetic data generated and analyzed during this study are stored on secure local File Transfer Protocol (FTP) servers at the Centre for Genomics and Oncological Research, Granada, Spain, research center and are available, upon reasonable request, from the corresponding author, Juan Sainz (jsainz@ugr.es). The functional data used in this project were meticulously catalogued and archived in the Biobanking and BioMolecular resources Research Infrastructure – Netherlands (BBMRI-NL) data infrastructure (<https://hfgp.bbmri.nl/>) using the Molgenis open-source platform for

scientific data. This allows flexible data querying and download, including sufficiently rich metadata and interfaces for machine processing (R statistics, Representational State Transfer Application Programming Interface[REST API]), and Findable, Accessible, Interoperable, and Reusable (FAIR) principles to optimize the ability, accessibility, interoperability, and reusability.

The full-text version of this article contains a data supplement.

Licensed under [Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International \(CC BY-NC-ND 4.0\)](https://creativecommons.org/licenses/by-nc-nd/4.0/), permitting only noncommercial, nonderivative use with attribution.

Key Points

- CDKN2A and BCL2 SNPs are newly linked to CLL risk, with CDKN2A rs3731204 showing the strongest association ($P = 1.57 \times 10^{-12}$).
- These variants are linked to altered gene expression (eg, CDKN2A, ACTA2) and affect immune responses but not survival or treatment timing.

We investigated the influence of 55 583 autophagy-related single-nucleotide polymorphisms (SNPs) on chronic lymphocytic leukemia (CLL) risk across 4 independent populations comprising 5472 CLL cases and 726 465 controls. We also examined their impact on overall survival (OS), time to first treatment (TTFT), autophagy flux, and immune responses. A meta-analysis of the 4 populations identified, to our knowledge, for the first time, significant associations between *CDKN2A* (rs3731204) and *BCL2* (rs4940571, rs12457371, and rs1026825) SNPs and CLL risk, with *CDKN2A* showing the strongest association ($P = 1.57 \times 10^{-12}$). We also validated previously reported associations for *FAS*, *BCL2*, and *BAK1* SNPs with CLL risk ($P = 4.73 \times 10^{-21}$ to 3.39×10^{-9}). The *CDKN2A*_{rs3731204} and *FAS*_{rs1926194} SNPs associated with increased *CDKN2A* and *ACTA2* messenger RNA expression levels in the whole blood and/or lymphocytes ($P = 5.1 \times 10^{-7}$, $P = 1.58 \times 10^{-21}$, and $P = 7.8 \times 10^{-41}$), although no significant effect on autophagy flux was observed. However, associations were found between *CDKN2A*, *BCL2*, and *FAS* SNPs and various T-cell subsets, cytokine production, and circulating concentrations of interferon gamma, tumor necrosis factor-related apoptosis-inducing ligand, CD40, chemokine ligand 20, and interleukin-2 receptor subunit β proteins ($P \leq .005$). No significant association was detected between autophagy variants and OS or TTFT, suggesting that these variants drive disease initiation rather than progression. In conclusion, this study identified 4 novel associations for CLL and provided insights into the biological pathways that influence CLL development.

Introduction

Chronic lymphocytic leukemia (CLL) is an incurable disease¹ that accounts for 25% to 35% of new leukemia cases in Europe and the United States.^{2,3} Although occasionally diagnosed in younger individuals (aged <55 years),⁴ the median age at diagnosis is 71 years. The advent of Bruton tyrosine kinase and B-cell lymphoma 2 (*BCL2*) inhibitors has improved the 5-year survival rate to ~86%.⁵ Identified risk factors include age, sex, chemical exposure, European ancestry, and family history of hematological cancers.⁶ Prognostic factors, such as Rai and Binet staging, lymphocyte doubling time, CD38 and ZAP-70 expression, immunoglobulin mutation status, B-cell receptor stereotypy, cytogenetic changes, and gene mutations, are used in patient stratification.² Still, despite the success of genome-wide association studies (GWASs) in uncovering susceptibility and prognostic loci for CLL,⁷⁻¹¹ the etiopathogenesis of this heterogeneous disease remains poorly understood.

Recent studies implicate autophagy, a conserved catabolic process promoting cell survival, in CLL risk and prognosis.¹²⁻¹⁴ Autophagy contributes to malignant transformation and chemotherapeutic response, and has been observed in both hematological malignancies and solid tumors.⁷⁻¹¹ In CLL, abnormal expressions of autophagy-related genes (*BECN1*, *DAPK1*, *SLAMF1*, *GABARAPL2*, *ATG3*, *ATG5*, *ATG7*, and *TLR9*) is linked to disease progression.¹⁵⁻²¹ Several reports show that autophagy has a prosurvival role, and its inhibition induces programmed cell death in CLL.¹² These findings align with evidence of autophagy-apoptosis interplay in cancer and normal cells.^{22,23} However, a selective role of autophagy in CLL has been suggested, making it a

potential therapeutic target. Conversely, a study in acute myeloid leukemia using Unc-51-like autophagy activating kinase 1 knockout mice showed that autophagy induction helped overcome venetoclax resistance,²⁴ a standard CLL therapy. Moreover, autophagy activation associates with better outcomes in other hematological cancers,²⁵ suggesting a context-dependent function.

Given this, we investigated whether genetic variants in autophagy-related genes affect CLL risk across 4 European-ancestry populations. We also assessed their influence on time to first treatment (TTFT) and overall survival (OS). Lastly, considering autophagy's role in immune modulation,²⁶⁻²⁹ we explored associations between these variants and autophagy flux in 67 healthy individuals, as well as immune traits in the 500FG cohort of the Human Functional Genomic Project (HFGP).³⁰⁻³²

Materials and methods

Discovery populations

The discovery populations included 4272 patients with CLL and 724 567 healthy controls from the International Lymphoma Epidemiology Consortium (InterLymph; 3099 patients, 7666 controls), the FinnGen project (668 patients, 314 189 controls), and the UK Biobank (505 patients, 402 712 controls). CLL cases were diagnosed according to updated international criteria,² and controls were randomly selected healthy individuals. The InterLymph cohort comprised 4 previous GWASs: National Cancer Institute (NCI) non-Hodgkin lymphoma (NHL) GWAS,¹⁷ Utah CLL GWAS, Genetic Epidemiology of CLL Consortium GWAS,¹⁸ and University of California San Francisco Molecular Epidemiology of

NHL GWAS.¹⁹ Patients were recruited via clinics, hospitals, cancer registries, or self-reports confirmed by medical and pathology records.

The InterLymph Data Coordinating Center reviewed International Classification of Diseases (ICD) codes and pathology/medical data, classifying cases per the World Health Organization–based hierarchical system of the InterLymph Pathology Working Group (2008).³³ Genotyping for InterLymph GWASs was conducted using the Illumina OmniExpress platform at the NCI Cancer Genomic Research Laboratory. Genotypes were called with Illumina GenomeStudio, and duplicate concordance exceeded 99%. Quality control excluded monomorphic single-nucleotide polymorphisms (SNPs) and variants with call rate of <93%. Samples were removed if they had call rate of $\leq 93\%$, autosomal heterozygosity of <0.25 or >0.33, or sex discordance (>5% X chromosome heterozygosity in males, <20% in females). Unexpected duplicates (>99.9% concordance) and first-degree relatives (Pihat of >0.40) were also excluded. Ancestry was assessed via the genotype likelihoods (GLs) structure admix module following Pritchard et al,³⁴ excluding individuals with <80% European ancestry.

GWAS data from UK Biobank and FinnGen were retrieved from their repositories.^{35,36} End point definitions were based on electronic health records and ICD codes (UK Biobank), and nationwide registries. Further details have been described previously.^{35,36}

Selection of SNPs and meta-analysis of discovery cohorts

A total of 234 autophagy-related genes were selected from the autophagy database (<http://autophagy.lu/>, accessed in November 2023; supplemental Table 1). Association estimates for 55 583 genotyped or imputed SNPs located within or near these genes (5 kilobases upstream, 3 kilobases downstream) were extracted from the discovery GWAS data sets and meta-analyzed using METAL.³⁷ Genomic control was applied to correct for population stratification, and the I^2 statistic assessed heterogeneity across studies. Pooled odds ratios (ORs) were calculated using a fixed-effects model.

Of 55 583 variants, 49 612 SNPs were shared across all GWAS data sets and included in the association analysis. Among these, 1903 were considered common (minor allele frequency of > 0.05) and independent ($r^2 < 0.1$) based on LDLink data for European populations (<https://ldlink.nci.nih.gov/?tab=snpclip>). A multiple testing significance threshold was set at $P = 2.63 \times 10^{-5}$ (0.05/1903 SNPs; supplemental Table 2).

Replication cohort, genotyping, and meta-analysis

After meta-analyzing GWAS data from InterLymph, UK Biobank, and FinnGen, we selected independent autophagy-related SNPs associated with CLL risk based on the following criteria: $r^2 < 0.1$ (linkage disequilibrium threshold), $P \leq .0001$, and no significant heterogeneity across populations (heterozygosity $P \geq .05$). Previously reported CLL susceptibility SNPs were excluded. Eleven SNPs met these criteria and were advanced for replication in the CRuCIAL cohort, comprising 1200 patients with CLL and 1898 controls (supplemental Table 3).

The CRuCIAL study adhered to the Declaration of Helsinki and was approved by the institutional review boards of all participating centers. Details of the cohort are in supplemental Table 3. Genotyping was performed at the Centre for Genomics and Oncological Research, Granada, Spain, using KASPar assays (LGC Genomics, Middlesex, United Kingdom), following established protocols.³⁸ For quality control, ~5% of samples were genotyped in duplicate, showing $\geq 99.0\%$ concordance. Genotype frequencies matched those in the 1000 Genomes database and were in Hardy-Weinberg equilibrium.

Association analyses were conducted using STATA version 12.1, with statistical power estimated via Quanto version 12.4 (log-additive model). Analyses were adjusted for age, sex, and country of origin. Finally, we performed a meta-analysis combining CRuCIAL results with discovery cohorts using METAL,³⁷ as described earlier.

OS and TTFT analyses in the CRuCIAL cohort

To assess the impact of autophagy-related variants on patient survival and disease progression, we evaluated associations between the selected SNPs and either OS or TTFT in the CRuCIAL cohort. The primary outcome was OS, defined as time from CLL diagnosis to death from any cause, with censoring at death or last follow-up. TTFT was defined as the time from diagnosis to first treatment or last follow-up. Associations with OS and TTFT were analyzed using Cox regression models adjusted for age, sex, and country of origin. Hazard ratios (HRs) were estimated for each SNP. For susceptibility analyses, the significance threshold was $P = 2.63 \times 10^{-5}$ (0.05/1903 SNPs; supplemental Table 2). Analyses were conducted using STATA version 12.1 (StataCorp, College Station, TX), and statistical power was calculated with the survSNP package in R (version 4.1.1; R Core Team, 2018).

Impact of autophagy-related variants on autophagy flux

We investigated whether autophagy-related SNPs were associated with autophagy flux in 68 healthy donors of European ancestry. Peripheral blood mononuclear cells (PBMCs) were isolated by density gradient centrifugation using Histopaque and treated for 2 hours with 10 μM bafilomycin A1 (inhibitor) or 10 mM metformin (inducer). A total of 5×10^5 PBMCs were plated per well in 96-well round-bottom plates (Greiner) and treated with either metformin or bafilomycin A1, both, or left untreated. After treatment, cells were lysed in 50 μL of buffer (1% NP-40, 500 mM Tris HCl, 2.5 M NaCl, 20 mM EDTA, and phosphatase and protease inhibitors; pH 7.2). Twenty micrograms of protein were resolved by 12% sodium dodecyl sulfate–polyacrylamide gel electrophoresis, transferred to nitrocellulose membranes (Trans-Blot Turbo), blocked in Tris-buffered saline with Tween 20 with 5% bovine serum albumin, and incubated overnight at 4°C with primary antibodies (rabbit anti-LC3A/B, Cell Signaling; mouse anti-actin, Merck Millipore) diluted 1:1000 in 1% bovine serum albumin. After washing, secondary antibodies were applied (anti-rabbit immunoglobulin G or anti-mouse immunoglobulin G) were applied. Detection was performed using SuperSignal West Femto (Thermo Fisher) or Clarity Western ECL (Bio-Rad), and images were captured with a ChemiDoc XRS system. Densitometry was

done using Quantity One version 4.6.5 (Bio-Rad). Autophagy synthesis was calculated as the LC3-II ratio between metformin and Baf A1 and Baf A1-only conditions; degradation was assessed by comparing metformin with/without Baf A1. Analyses followed standard autophagy guidelines.³⁹ Linear regression adjusted for age and sex tested SNP-flux associations, with a significance set at $P = .01$ (5 SNPs, 2 treatments).

Functional effect of the autophagy-related variants on immune responses

To explore the functional relevance of SNPs that remained significant after correction for multiple testing ($P = 2.63 \times 10^{-5}$), as well as known CLL susceptibility loci, we assessed their association with cytokine expression quantitative trait loci (cQTL) data from in vitro stimulation experiments in 408 healthy individuals from the 500FG cohort of the HFGP.

cQTL data included levels of interferon gamma (IFN- γ), interleukin-1 β (IL-1 β), IL-6, tumor necrosis factor α , IL-17, and IL-22 measured in PBMCs, monocyte-derived macrophages, or whole blood after 24-hour stimulation with lipopolysaccharide (1 or 100 ng/mL; Sigma-Aldrich), phytohemagglutinin (10 μ g/mL; Sigma), N-palmitoyl-S-[2,3-bis(palmitoyloxy)-(R)-propyl]-cysteine (Pam3Cys; 10 μ g/mL; EMC Microcollections), or cytosine-phosphate-guanine (100 ng/mL; InvivoGen), or in unstimulated conditions. Cytokine concentrations were determined using commercial enzyme-linked immunosorbent assay kits (PeliKine Compact or R&D Systems), following manufacturer instructions. For values outside detection limits, the respective limit was assigned. After log transformation, linear regression analyses adjusted for age and sex were used to assess associations between selected SNPs and cytokine levels. Detailed protocols for PBMC isolation, macrophage differentiation, and stimulation assays have been described previously.^{31,40}

Association of autophagy SNPs and blood cell counts and serum/plasma proteomic profiles

To assess the impact of selected SNPs on immune cell variation, 91 blood-derived cell populations were quantified by 10-color flow cytometry (Navios, Beckman Coulter) within 2 to 3 hours of sampling. Cell populations were analyzed using Kaluza software (version 1.3), and interexperimental noise was minimized by calculating parental and grandparental percentages, that is, the proportion of a given cell type within its immediate or ancestral subset (supplemental Table 4). Detailed protocols for cell isolation, reagents, gating, and flow cytometry have been published elsewhere.⁴¹

Proteomic profiling was also performed on serum and plasma samples from the HFGP cohort using the Olink inflammation panel (Olink, Sweden), which quantified 103 circulating proteins (supplemental Table 5). Protein levels were log₂ transformed and normalized using bridging samples to adjust for batch effects.

To correct for multiple testing, significance thresholds were set at 2.92×10^{-4} for cQTL, 2.55×10^{-5} for proteomic, and 2.89×10^{-5} for cell count analyses. Methodological details for all functional assays are available in previous publications.^{30-32,40} Analyses were performed in R (<http://www.r-project.org>) using custom scripts, and plots were generated with Prism.

In silico functional analysis

We used Haploreg (<http://www.broadinstitute.org/mammals/haploreg/haploreg.php>) to predict the functional roles of autophagy-related SNPs. Their potential as expression QTL across tissues was assessed using genotype-tissue expression project (GTEx; <https://gtexportal.org/home/>), and broader QTL features were explored via QTLbase (<http://www.mulinlab.org/qtlbase>), which compiles data from GTEx, the Cancer Genome Atlas, Database of Immune Cell Expression, and others. To complement these analyses, meta-scores integrating conservation, allele frequency, and predicted protein impact were generated. Additional functional evaluation included combined annotation-dependent depletion,⁴² RegulomeDB (<https://regulomedb.org/>), and FORGEdb (<https://forge2.altiusinstitute.org/files/forgefdb.html>).

Ethics approval and consent to participate

Ethical approval was obtained from each participating institution for the InterLymph, UK Biobank, and FinnGen GWASs. In addition, the CRuCIAL study was approved by the ethical review committee of participant institutions: Virgen de las Nieves University Hospital (Granada, Spain, 0760-N-18); University Hospital of Salamanca (Salamanca, Spain, PI90/07/2018); Hospital del Mar (Barcelona, Spain); Catalan Institute of Oncology (Barcelona, Spain); Morales Meseguer University Hospital (Murcia, Spain); Consortium for Biomedical Research in Epidemiology and Public Health (CIBERESP) group (Spain); University of Modena and Reggio Emilia, AOU Policlinico (Modena, Italy); University of Pisa (Pisa, Italy), Wroclaw Medical University (Wroclaw, Poland); and the Radboud University Medical Center (Nijmegen, The Netherlands, 2011/299). A detailed description of the study cohort has been reported elsewhere.² All participants provided written informed consent to participate in these genetic studies. The HFGP study was approved by the Arnhem-Nijmegen ethical committee (no. 42561.091.12) and biological specimens were collected after informed consent was obtained.

Results

This study followed a 2-phase design combining discovery and validation stages. In the discovery phase, we performed a meta-analysis of 3 large European cohorts (InterLymph, UK Biobank, and FinnGen), including 4272 CLL cases and 724 567 healthy controls. This analysis identified multiple SNPs significantly associated with CLL risk. The strongest associations were observed for the *FAS*_{rs1926194}, *BAK1*_{rs210143}, and *BCL2*_{rs4987852} SNPs (OR = 1.29, $P = 4.73 \times 10^{-21}$; OR = 1.20, $P = 3.39 \times 10^{-9}$; and OR = 1.38, $P = 5.40 \times 10^{-12}$, respectively), which served as internal positive controls and remained statistically significant in the combined analysis. In addition to these previously reported loci, another 11 independent variants including *BCL2*_{rs1026825}, *BCL2*_{rs11152374}, *BCL2*_{rs12457371}, *BCL2*_{rs4940571}, *BCL2*_{rs7236090}, *CDKN2A*_{rs3731204}, *EGFR*_{rs4947976}, *LPAR6|RB1*_{rs11839271}, *NAF1*_{rs6829366}, *PRKCD*_{rs143052840}, and *TFEB*_{rs6910366} also showed notable associations and were selected for replication based on their statistical significance, effect sizes, independence, and low between-study heterogeneity (Table 1).

In the replication phase, these 11 SNPs were tested in an independent cohort (CRuCIAL), consisting of 1200 CLL cases and 1898 controls. Interestingly, the *CDKN2A*_{rs3731204} and

Table 1. Meta-analysis of the autophagy-related risk variants for CLL in the discovery cohorts

SNP	Gene	Chr	Effect allele	InterLymph cohort (N = 3099 CLL cases and 7666 healthy controls)		UK Biobank (N = 505 CLL cases and 402 712 healthy controls)		FinnGen Risteys 10 (N = 668 CLL cases 314 189 healthy controls)		Discovery meta-analysis (N = 4272 CLL cases and 724 567 healthy controls)		
				OR (95% CI)	P value	OR (95% CI)	P value	OR (95% CI)	P value	OR (95% CI)	P value	<i>phet</i>
rs59952010	<i>ARSB</i>	5	T	1.13 (1.06-1.21)	2.16 × 10 ⁻⁴	1.17 (1.03-1.34)	.020	1.05 (0.93-1.19)	.418	1.13 (1.06-1.19)	6.29 × 10 ⁻⁵	0.831
rs210143	<i>BAK1</i>	6	C	1.21 (1.13-1.30)	9.52 × 10⁻⁸	1.08 (0.94-1.24)	.260	1.29 (1.13-1.48)	1.65 × 10 ⁻⁴	1.20 (1.13-1.27)	3.39 × 10⁻⁹	0.083
rs4987852	<i>BCL2</i>	18	C	1.43 (1.28-1.61)	1.61 × 10⁻⁹	1.06 (0.84-1.34)	.640	1.47 (1.27-1.72)	5.30 × 10⁻⁷	1.38 (1.26-1.51)	5.40 × 10⁻¹²	0.148
rs4987856	<i>BCL2</i>	18	T	0.69 (0.66-0.77)	3.55 × 10⁻¹¹	0.75 (0.60-0.93)	7.50 × 10 ⁻³	0.87 (0.68-1.11)	.249	0.72 (0.66-0.80)	7.35 × 10⁻¹¹	0.518
rs1026825	<i>BCL2</i>	18	G	1.09 (1.03-1.16)	4.08 × 10 ⁻³	1.21 (1.07-1.37)	3.50 × 10 ⁻³	1.10 (0.99-1.23)	.078	1.12 (1.06-1.18)	6.62 × 10 ⁻⁵	0.672
rs11152374	<i>BCL2</i>	18	A	1.12 (1.04-1.21)	3.72 × 10 ⁻³	1.15 (0.99-1.34)	.078	1.09 (0.97-1.24)	.159	1.12 (1.05-1.20)	5.42 × 10 ⁻⁴	0.377
rs4940571	<i>BCL2</i>	18	G	1.20 (1.08-1.34)	9.11 × 10 ⁻⁴	1.19 (0.96-1.47)	.120	1.13 (0.96-1.34)	.143	1.18 (1.08-1.29)	3.05 × 10 ⁻⁴	0.969
rs7236090	<i>BCL2</i>	18	C	1.10 (1.03-1.17)	3.84 × 10 ⁻³	1.16 (1.02-1.32)	.017	1.09 (0.98-1.22)	.121	1.11 (1.05-1.17)	1.92 × 10 ⁻⁴	0.896
rs12457371	<i>BCL2</i>	18	G	0.89 (0.83-0.94)	3.66 × 10 ⁻⁴	0.91 (0.79-1.05)	.210	0.86 (0.77-0.97)	.010	0.88 (0.83-0.93)	2.77 × 10 ⁻⁵	0.988
rs3731204	<i>CDKN2A</i>	9	C	0.79 (0.72-0.87)	4.42 × 10⁻⁷	0.76 (0.64-0.91)	2.90 × 10 ⁻³	0.76 (0.63-0.91)	3.41 × 10 ⁻³	0.78 (0.72-0.85)	1.03 × 10⁻⁹	0.972
rs4947976	<i>EGFR</i>	7	A	1.03 (0.97-1.09)	.383	1.22 (1.08-1.38)	2.40 × 10 ⁻³	1.12 (1.01-1.25)	.039	1.15 (1.07-1.24)	1.15 × 10 ⁻⁴	0.780
rs1926194	<i>FAS</i>	10	A	1.27 (1.19-1.35)	1.15 × 10⁻¹³	1.32 (1.16-1.50)	1.70 × 10⁻⁵	1.34 (1.20-1.50)	1.61 × 10⁻⁷	1.29 (1.23-1.36)	4.73 × 10⁻²¹	0.145
rs7584971	<i>ITGA6</i>	2	A	1.01 (0.93-1.11)	.800	1.20 (1.00-1.44)	.050	1.22 (1.08-1.38)	1.14 × 10 ⁻³	1.17 (1.07-1.29)	6.07 × 10 ⁻⁴	0.578
rs11839271	<i>LPAR6, RB1</i>	13	T	0.90 (0.84-0.96)	2.28 × 10 ⁻³	0.92 (0.80-1.05)	.220	0.89 (0.79-1.00)	.052	0.90 (0.85-0.96)	4.34 × 10 ⁻⁴	0.991
rs6829366	<i>NAF1</i>	4	T	1.13 (1.05-1.21)	1.49 × 10 ⁻³	1.16 (1.00-1.35)	.055	1.05 (0.91-1.21)	.478	1.12 (1.05-1.20)	4.26 × 10 ⁻⁴	0.022
rs143052840	<i>PRKCD</i>	3	A	1.25 (1.09-1.44)	1.14 × 10 ⁻³	1.54 (1.19-1.98)	7.30 × 10 ⁻⁴	0.96 (0.72-1.29)	.786	1.26 (1.11-1.42)	1.80 × 10 ⁻⁴	0.339
rs6910366	<i>TFEB</i>	6	G	0.83 (0.76-0.91)	5.11 × 10 ⁻⁵	0.84 (0.70-1.02)	.068	0.94 (0.81-1.09)	.429	0.86 (0.80-0.93)	8.28 × 10 ⁻⁵	0.850
rs2645488	<i>VMP1</i>	17	G	1.14 (1.07-1.22)	1.60 × 10 ⁻⁴	1.19 (1.03-1.36)	.017	0.95 (0.85-1.07)	.403	1.11 (1.04-1.18)	7.71 × 10 ⁻⁴	0.043
rs17885803	<i>WRAP53, TP53</i>	17	T	1.27 (1.11-1.44)	2.93 × 10 ⁻⁴	1.36 (1.06-1.76)	.017	1.10 (0.93-1.31)	.265	1.22 (1.11-1.36)	1.10 × 10 ⁻⁴	0.480

Estimates calculated according to a log-additive model of inheritance and adjusted for age, sex and country of origin. The meta-analysis was performed assuming a fixed-effect model using METAL. $P < 2.63E-5$ in bold. Chr, chromosome.

*BCL2*_{rs4940571} SNPs showed consistent directions of effect compared with the discovery phase and replicated with nominal significance (OR = 0.80, $P = 8.72 \times 10^{-3}$; OR = 1.40, $P = 8.66 \times 10^{-4}$, respectively), whereas some SNPs such as *BCL2*_{rs1026825} (OR = 1.06, $P = .353$) and *BCL2*_{rs12457371} (OR = 0.92, $P = .294$) showed the same direction but did not reach statistical significance (Table 1).

The meta-analysis of these 4 independent populations confirmed novel and statistically significant associations of *CDKN2A*_{rs3731204}, *BCL2*_{rs1026825}, *BCL2*_{rs12457371}, and *BCL2*_{rs4940571} SNPs with CLL risk (OR = 0.78; 95% confidence interval [CI], 0.73-0.84; $P = 1.57 \times 10^{-12}$; OR = 1.11; 95% CI, 1.06-1.16; $P = 1.75 \times 10^{-5}$; OR = 0.89; 95% CI, 0.84-0.93; $P = 3.69 \times 10^{-6}$; and OR = 1.21; 95% CI, 1.12-1.31; $P = 3.25 \times 10^{-6}$; respectively; Table 2).

The most statistically significant effect on CLL risk was found for *CDKN2A*_{rs3731204} SNP within the *CDKN2A* locus. Each copy of *CDKN2A*_{rs3731204C} allele decreased the risk of developing CLL by 22%. According to the GTEx data, the *CDKN2A*_{rs3731204C} allele was strongly associated with higher levels of *CDKN2A* messenger RNA (mRNA) expression levels in whole blood ($P = 5.1 \times 10^{-7}$; supplemental Figure 1A). In addition, although this SNP was not significantly associated with autophagy flux levels ($P > .01$), our functional experiments revealed, to our knowledge, for the first time, an association between the *CDKN2A*_{rs3731204C} protective allele and decreased absolute numbers of CD45RO-CD45RA⁺ effector T cells and CD4⁺CD8⁺ T cells ($P = .0027$ and $P = .0036$; Figure 1A-B). Moreover, we found that carriers of the *CDKN2A*_{rs3731204C} protective allele had increased levels of IFN- γ after stimulation of whole blood with phytohemagglutinin ($P = .0040$; Figure 1C), suggesting a functional role of this SNP in CLL risk through modulation of *CDKN2A* protein levels and specific subsets of immune cells.

In addition to the association of *CDKN2A* SNP with CLL risk, we found novel and statistically significant associations for *BCL2*_{rs1026825}, *BCL2*_{rs12457371}, and *BCL2*_{rs4940571} SNPs with CLL risk (Table 2). These associations were independent of those previously reported in this locus (rs4987852 and rs4987856). Intriguingly, we found no association between *BCL2* SNPs and autophagic flux levels. However, carriers of the *BCL2*_{rs4940571G} risk allele had increased absolute numbers of FOXP3⁺Helios⁺ regulatory T (Treg) cells ($P = .005$; Figure 2A) and decreased circulating concentrations of IFN- γ ($P = .0044$; Figure 2B). In contrast, we found that carriers of the *BCL2*_{rs1026825G} risk allele had increased absolute numbers of CD4⁺CD25⁺CD127^{low} Treg cells ($P = .0036$; Figure 2C) and decreased circulating concentrations of IL-2 receptor subunit β ($P = .0058$; Figure 2D). In addition, we found that carriers of the *BCL2*_{rs1026825G} risk allele had increased production of IL-6 after in vitro stimulation of PBMCs with Pam3Cys ($P = .0036$; Figure 2E). In silico analysis also revealed that this SNP is strongly associated with a chromatin accessibility QTL in lymphocytes ($P = 4.15 \times 10^{-7}$). Although none of the functional findings performed in the HFPG cohort were significant after correction for multiple testing, these results suggest that *BCL2*_{rs4940571} and *BCL2*_{rs1026825} SNPs might influence CLL risk by determining the absolute numbers of Treg cells, IL-6-mediated immune responses, and changes in chromatin accessibility.

In addition to these novel findings, the meta-analysis of the 4 European populations also validated previously reported associations with CLL risk for 4 genetic variants within *FAS*, *BCL2*, and *BAK1* (Table 2). As expected, all of these associations remained significant after correction for multiple testing. In GTEx, the *FAS*_{rs1926194A} risk allele is strongly associated with actin- $\alpha 2$ (*ACTA2*) mRNA expression levels in whole blood and lymphocytes ($P = 7.8 \times 10^{-41}$ and $P = 1.58 \times 10^{-21}$, respectively; supplemental Figure 1B), which suggest that this SNP might influence CLL risk through the modulation of gene expression of *ACTA2*, a well-known susceptibility locus for CLL. Interestingly, although we failed to find a significant association between this SNP and autophagy flux levels ($P > .01$; supplemental Table 6), we found in the HFPG that the healthy carriers of the *FAS*_{rs1926194A} risk allele had increased circulating concentrations of tumor necrosis factor-related apoptosis-inducing ligand (TRAIL), chemokine ligand 20, and CD40 ($P = .0036$, $P = .0018$, and $P = .0039$, respectively; Figure 3). Although none of these functional findings remained significant after multiple testing corrections, they suggested that this genetic variant may modulate the risk of CLL through host immune-mediated mechanisms. No functional effect for the *BCL2*_{rs4987852}, *BCL2*_{rs4987856}, and *BAK1*_{rs210143} SNPs was found in relation to autophagy flux or immune responses, suggesting that the effect of these SNPs on CLL risk is not mediated by these mechanisms.

Finally, given the noticeable impact of autophagy-related SNPs on CLL risk and the known role of *CDKN2A* and *BCL2* in determining tumor progression and drug response in many cancer types,¹ we tested whether selected autophagy-related SNPs could influence the OS of patients with CLL and TTFT in the CRuCIAL cohort. Our results showed no association between autophagy-related variants and either OS or TTFT (supplemental Tables 7 and 8). We only found a modest association of the *TFEB*_{rs6910366} SNP with longer OS (HR, 0.70; 95% CI, 0.52-0.94; $P = .016$) and an association of the *BCL2*_{rs4940571} SNP with shorter TTFT (HR, 1.43; 95% CI, 1.01-2.04; $P = .046$). Neither association remained significant after multiple testing corrections, suggesting that autophagy-related SNPs do not have a strong effect on patient survival or disease progression.

Discussion

This comprehensive study found, to our knowledge, for the first time that the *CDKN2A* locus is a susceptibility gene for CLL. In addition, this study identified 3 new susceptibility SNPs within *BCL2* that were independent of those previously reported at this locus. These results confirm the important role of *BCL2* in modulating CLL risk.

The most significant effect on CLL risk was found for *CDKN2A*_{rs3731204} SNP, which lies in the second intron of *CDKN2A*, a tumor suppressor gene that regulates key biological processes such as B-cell division, cell differentiation, autophagy, and cell death. *CDKN2A* encodes a cell-cycle inhibitor protein (p16^{*CDKN2A*}), which impedes abnormal cell growth and proliferation by binding to complexes of cyclin-dependent kinases (CDK) 4 and 6, and cyclin D, which leads to cell cycle arrest in the G1/S phase.⁴³ Both common and rare germ line variants or deletions within *CDKN2A* have been consistently associated with

Table 2. Overall meta-analysis of association estimates of autophagy-related risk variants for CLL

SNP	Gene	Chr	Effect allele	Discovery meta-analysis			CRuCIAL cohort		Overall meta-analysis			References (PMID)
				(N = 724 567 healthy controls; 4272 CLL cases)			(N = 1898 healthy controls; 1200 CLL cases)		(N = 726 465 healthy controls; 5472 CLL cases)			
				OR (95% CI)	P value	PHet	OR (95% CI)	P value	OR (95% CI)	P value	PHet	
rs59952010	ARSB	5	T	1.13 (1.06-1.19)	6.29E-05	0.831	–	–	–	–	–	26956414
rs210143	BAK1	6	C	1.20 (1.13-1.27)	3.39E-09	0.083	–	–	–	–	–	22700719; 24292274; 23770605; 28165464
rs4987852	BCL2	18	C	1.38 (1.26-1.51)	5.40E-12	0.148	–	–	–	–	–	23770605
rs4987856	BCL2	18	T	0.72 (0.66-0.80)	7.35E-11	0.518	–	–	–	–	–	26956414; 23770605
rs1026825	BCL2	18	G	1.12 (1.06-1.18)	6.62E-05	0.672	1.06 (0.94-1.19)	.353	1.11 (1.06-1.16)	1.75E-05	0.698	–
rs11152374	BCL2	18	A	1.12 (1.05-1.20)	5.42E-04	0.377	0.97 (0.84-1.13)	.716	1.10 (1.04-1.16)	1.13E-03	0.193	–
rs11839271	LPAR6, RB1	13	T	0.90 (0.85-0.96)	4.34E-04	0.991	1.13 (0.96-1.33)*	.131	0.92 (0.88-0.97)	1.63E-03	0.268	–
rs12457371	BCL2	18	G	0.88 (0.83-0.93)	2.77E-05	0.988	0.92 (0.78-1.08)	.294	0.89 (0.84-0.93)	3.69E-06	0.989	–
rs143052840	PRKCD	3	A	1.26 (1.11-1.42)	1.80E-04	0.339	0.82 (0.62-1.00)*	.152	1.18 (1.06-1.31)	1.59E-03	0.027	–
rs3731204	CDKN2A	9	C	0.78 (0.72-0.85)	1.03E-09	0.972	0.80 (0.67-0.94)	8.72E-03	0.78 (0.73-0.84)	1.57E-12	0.982	–
rs4940571	BCL2	18	G	1.18 (1.08-1.29)	3.05E-04	0.969	1.40 (1.15-1.71)	8.66E-04	1.21 (1.12-1.31)	3.25E-06	0.445	–
rs1926194	FAS	10	A	1.29 (1.23-1.36)	4.73E-21	0.145	–	–	–	–	–	24292274; 26956414; 28165464
rs7584971	ITGA6	2	A	1.17 (1.07-1.29)	6.07E-04	0.578	–	–	–	–	–	32887889
rs4947976	EGFR	7	A	1.15 (1.07-1.24)	1.15E-04	0.780	1.02 (0.91-1.14)†	.723	1.11 (1.05-1.17)	4.04E-04	0.360	–
rs6829366	NAF1	4	T	1.12 (1.05-1.20)	4.26E-04	0.022	1.14 (0.98-1.31)	.087	1.12 (1.06-1.19)	5.73E-05	0.024	–
rs6910366	TFEB	6	G	0.86 (0.80-0.93)	8.28E-05	0.850	1.07 (0.85-1.33)*	.574	0.87 (0.82-0.93)	8.74E-05	0.474	–
rs7236090	BCL2	18	C	1.11 (1.05-1.17)	1.92E-04	0.896	0.99 (0.88-1.12)	.912	1.10 (1.04-1.15)	1.83E-04	0.746	–
rs2645488	VMP1	17	G	1.11 (1.04-1.18)	7.71E-04	0.043	–	–	–	–	–	31407831
rs17885803	WRAP53, TP53	17	T	1.22 (1.11-1.36)	1.10E-04	0.480	–	–	–	–	–	32887889

Estimates calculated according to a log-additive model of inheritance and adjusted for age and sex. Meta-analysis was performed assuming a fixed-effect model using METAL software. $P < 2.63E-05$ in bold. Chr, Chromosome; PHet, P value for heterogeneity.

*Association estimates was calculated in a subset of 1 055 healthy controls and 1 024 CLL cases from the CRuCIAL cohort.

†Authors report the effect found for the rs12718945 (an SNP in complete linkage disequilibrium with the rs4947976, $r^2 = 0.996$).

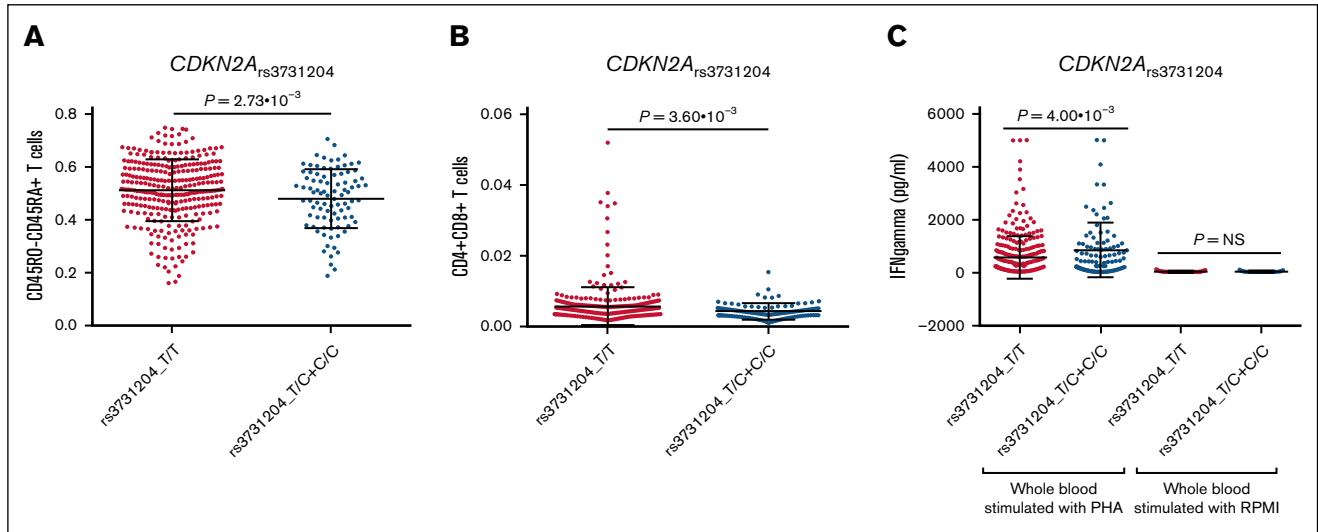


Figure 1. Functional impact of the *CDKN2A*_{rs3731204} SNP. Association of the *CDKN2A*_{rs3731204} SNP with absolute numbers of specific subsets of T cells (A and B) and IFN- γ levels after in vitro stimulation of WB with PHA (C). NS, nonsignificant; PHA, phytohemagglutinin; WB, whole blood.

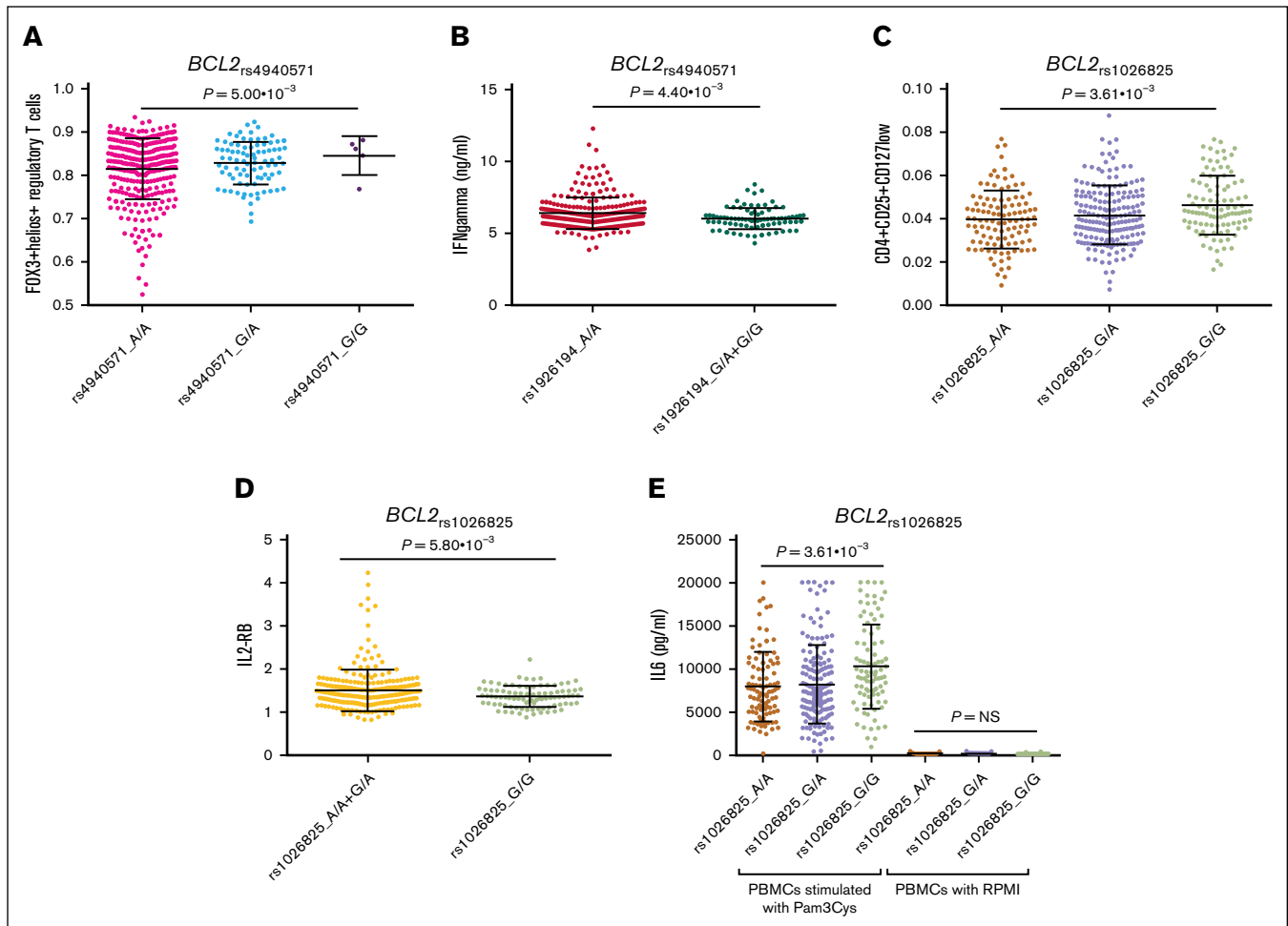


Figure 2. Functional impact of the *BCL2*_{rs1026825} and *BCL2*_{rs4940571} SNPs. Association of the *BCL2*_{rs1026825} and *BCL2*_{rs4940571} SNPs with absolute numbers of specific subsets of T cells and regulatory T cells, circulating concentrations of inflammatory proteins (IFN- γ and IL-2 receptor subunit β) and IL-6 levels after stimulation of PBMCs with Pam3Cys. NS, nonsignificant; Pam3Cys, N-palmitoyl-S-[2,3-bis(palmitoyloxy)-(R)-propyl]-cysteine.

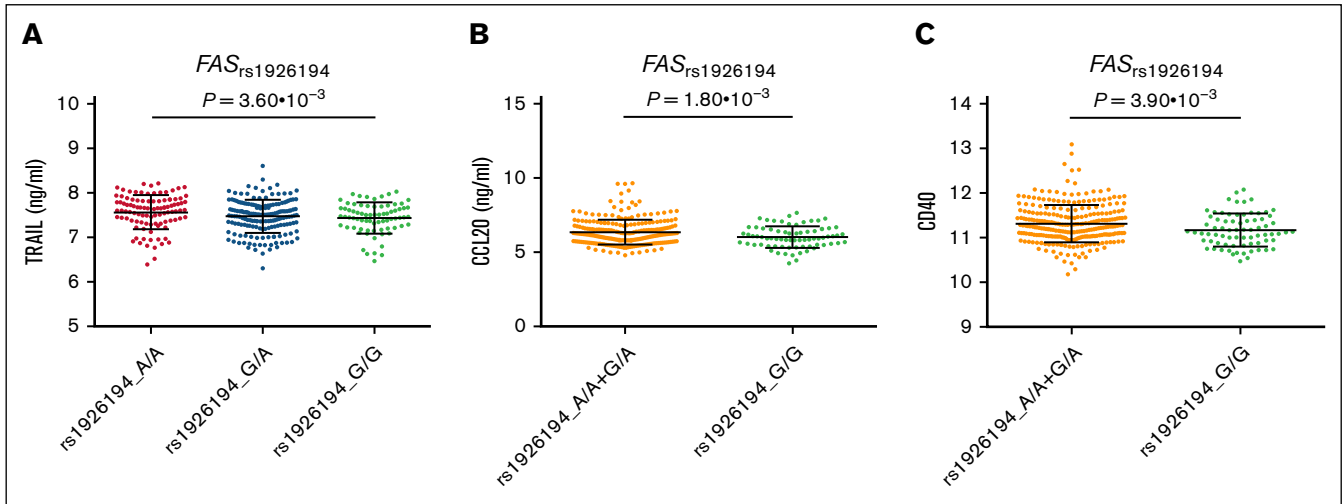


Figure 3. Functional impact of *FAS*_{rs1926194} SNP. Association of the *FAS*_{rs1926194} SNP with circulating concentrations of TRAIL, CCL20, and CD40. CCL20, chemokine ligand 20.

melanoma⁴⁴ and endocrine pancreatic cancer,⁴⁵ as well as blood cancers such as multiple myeloma,⁴⁰ acute lymphocytic leukemia,⁴⁶ and diffuse large B-cell lymphoma.⁴⁷

In this study, we found that carriers of the *CDKN2A*_{rs3731204C} allele (associated with higher *CDKN2A* mRNA expression levels according to the GTEx data) had a decreased risk of developing CLL. This finding agreed with previous results, suggesting that the loss of function of *CDKN2A* might lead to autophagy deregulation⁴⁸ and cancer development through inactivation of destabilization of the p53 protein, which increases cell division rates and inhibits p53-mediated apoptosis. In line with this hypothesis, Haploreg and QTLbase data showed that the *CDKN2A*_{rs3731204} SNP maps on histone marks (H3K4me1 and H3K4me3) in multiple immune cell types and alters binding sites for key transcription factors such as DMRT1-7, HDAC2, FOXD3, and Evi1, which play important roles in epigenetic repression, chromatin structure and transcriptional regulation, cell cycle progression, developmental events, and CLL progression. In addition, we found that healthy individuals carrying the *CDKN2A*_{rs3731204C} protective allele had decreased numbers of CD45RO⁺CD45RA⁺ naïve T cells in the blood, which were found to be enriched in patients with CLL with advanced Rai stages (III or IV).⁴⁹ In addition, it has been observed that these cells are unresponsive to tumor antigens, which might indicate that these cells act as drivers of disease onset and progression. In contrast, we observed that carriers of the *CDKN2A*_{rs3731204C} allele had decreased numbers of double CD4⁺CD8⁺ T cells, which are a type of T cell frequently found in patients with CLL with an active disease.⁵⁰ Although no definitive proof exists, it has been proposed that these cells are involved in CLL progression.⁵¹ In agreement with the hypothesis suggesting a role of the *CDKN2A* gene in determining host immunity, previous studies have reported that the *CDKN2A* gene shapes T cell- and macrophage-mediated immune responses in different types of cancer⁵²⁻⁵⁴ and even mediates responses to immunotherapies.^{55,56} Given that we did not find a significant association between the *CDKN2A*_{rs3731204} SNP and autophagy flux levels, it seems plausible that this SNP influences the risk of CLL by

modulating the absolute numbers of specific T-cell subsets rather than through the control of the autophagy pathway.

Another interesting result was the association between CLL risk and 3 independent SNPs within the *BCL2* gene. The *BCL2* locus is a well-known susceptibility gene for CLL that encodes a protein that regulates apoptosis. Intriguingly, in contrast to many other oncogenes, *BCL2* does not promote cell proliferation but induces cell survival.⁵⁷ Although t(14;18) translocation affecting *BCL2* has been associated with the development of B-cell leukemias and lymphomas, multiple studies have demonstrated that the high expression of *BCL2* mRNA observed in most patients with CLL^{58,59} is not strictly dependent on t14;18 translocation.⁶⁰ Interestingly, the newly identified variants within the *BCL2* gene (rs1026825, rs12457371, and rs4940571) were independent of each other ($r^2 < 0.1$) and of those previously reported within this locus (rs4987852 and rs4987855),⁶¹ which confirmed the importance of *BCL2* in modulating the risk of CLL but also the difficulties in deciphering its complex genomic regulation and its role in CLL. Although multiple mechanisms have been proposed, including promoter hypomethylation,⁶² loss of microRNA-15 and microRNA-16 expression,⁶³ overexpression of nucleolin,⁶⁴ or even the existence of a 5'-untranslated region open reading frame that might repress *BCL2* mRNA expression,⁶⁵ the mechanism that drives *BCL2* expression in CLL remains unknown to date. Our functional experiments revealed no significant effect of *BCL2* SNPs on modulating autophagic flux levels. However, we found that carriers of the *BCL2*_{rs4940571G} risk allele had increased absolute numbers of FOXP3⁺Helios⁺ Treg cells, which are involved in suppressing the antitumor activity of multiple immune cell types, including effector and Treg cells, B cells, natural killer cells, natural killer T cells, monocytes, and dendritic cells. Several suppressive characteristics have been described for these Treg cells, including increased production of anti-inflammatory cytokines; enhanced release of perforins and granzymes; high expression of programmed cell death protein 1, cytotoxic T-lymphocyte-associated protein 4, and lymphocyte-activation gene 3; and interference in metabolism through IL-2 deprivation.^{66,67}

Furthermore, we observed that carriers of the *BCL2*_{rs4940571G} allele had decreased circulating concentrations of IFN- γ , which is a cytokine that activates the function of macrophages and cytotoxic T cells, and is also involved in modulating resistance to apoptosis⁶⁸ and CLL survival^{68,69} as well as the response to drugs used to treat patients with CLL.⁷⁰ In addition, we found that carriers of the *BCL2*_{rs1026825G} risk allele had increased numbers of CD4⁺CD25⁺CD127^{low} Treg cells, which increased with age⁷¹ and had specific regulatory and proliferative function in humans.⁷² Although none of these functional results remained significant after correction for multiple testing, in silico functional analysis also showed that *BCL2*_{rs12457371} and *BCL2*_{rs1026825} SNPs maps among histone marks, QTLs, and influence chromatin states in blood cells, which were proposed to play a role in the development of CLL.^{73,74} Additionally, we found that the *BCL2*_{rs1026825} SNP was strongly associated with a chromatin accessibility QT locus in lymphocytes, suggesting that this marker regulates the accessibility of transcription factors to chromatin and transcription. These results, along with those showing that the *BCL2*_{rs4940571} SNP alters the binding sites for B double prime 1, a transcription factor involved in RNA polymerase III transcription, cancer development, and tumor progression,⁷⁵ suggest that these SNPs might modulate the risk of CLL through immune- and nonimmune-related mechanisms.

This study also validated previously reported associations between SNPs within the *BCL2*, fas cell surface death receptor (*FAS*), and *BAK1* loci and CLL risk. Although none of these SNPs were associated with autophagy flux levels, our functional analyses revealed that healthy carriers of the *FAS*_{rs1926194A} risk allele had increased circulating concentrations of TRAIL and CD40. In line with these findings, it has been reported that TRAIL acts with FAS-ligand to synergistically induce apoptosis of CD40-activated CLL B cells,⁷⁶ and TRAIL also interacts also with “decoy” receptors (TRAIL-R3 and R4) to protect cells from apoptosis.⁷⁷ Furthermore, other authors have demonstrated that CLL cells are resistant to TRAIL-induced apoptosis⁷⁸ and that CD40 is involved in instructing CLL cells to attract monocytes via the C-C chemokine receptor type 2 pathway.⁷⁹ In addition, the *FAS*_{rs1926194} SNP was reported to be strongly associated with *ACTA2* mRNA expression levels in the whole blood and lymphocytes, which is a known susceptibility locus for CLL involved in cell migration^{80,81} and cancer development.⁸¹ These findings suggest that the *FAS*_{rs1926194} SNP might modulate CLL risk through the induction of an antiapoptotic effect of TRAIL on CLL cells and monocyte/macrophage recruitment, as well as by promoting cell migration and proliferation. In line with this hypothesis, we also found that carriers of the *FAS*_{rs1926194A} allele had increased circulating concentrations of chemokine ligand 20, a chemokine produced by monocytes and macrophages and involved in promoting cell migration, proliferation, metastasis, and cancer progression.^{82,83}

Although it was tempting to speculate that autophagy might be the mechanism affecting the risk of developing CLL, the markers in the *CDKN2A*, *BCL2*, *FAS*, and *BAK1* genes did not associate with autophagy flux levels. This confirms that these genes might not influence disease onset through the control of this mechanism, but rather through the modulation of multiple immunological and nonimmunological mechanisms, implicating specific immune cells, cytokines, and multiple transcription factors.

Finally, in this study, we evaluated whether autophagy-related variants were associated with OS and TTFT in the CRuCiAL cohort. Although we found a modest association between the *TFEB*_{rs6910366} and *BCL2*_{rs4940571} SNPs and longer OS and shorter TTFT, respectively, these associations did not remain significant after correction for multiple comparisons. Therefore, we concluded that autophagy-related SNPs do not have a significant effect on patient survival or disease progression. Although a previous study suggested that a promoter variant within *BCL2* was weakly associated with disease progression,⁸⁴ these results could not be confirmed by others,⁸⁵ which reinforced the idea of a null effect of autophagy variants on OS and disease progression.⁸⁶

The major strengths of our study are the comprehensive analysis of inherited genetic variation in 234 autophagy-related genes reported in the autophagy database and the inclusion of 4 large independent European populations, including 5472 CLL cases and 726 465 healthy controls. Additionally, we analyzed the impact of autophagy-related SNPs on the modulation of blood cell counts, serum and plasma metabolites, and host immune responses in a large cohort of healthy donors from the HFGP. We had >80% power (log-additive model) to detect an OR of 1.15 at $\alpha = 2.63 \times 10^{-5}$ (multiple testing threshold) for a polymorphism with a minor allele frequency of at least 0.25. A weakness of this study was its multicentric nature, which placed inevitable drawbacks, such as the impossibility of collecting clinical history for a significant proportion of the patients analyzed. Finally, because all study participants included in this analysis were of European ancestry, we could not determine the impact of autophagy variants on CLL risk in other ethnic groups. Additional studies using other populations are warranted to confirm the results of this study.

Conclusion

In conclusion, this study identified, to our knowledge, for the first time, *CDKN2A* as a new susceptibility locus for CLL. Importantly, this study also identified new susceptibility variants for *BCL2* and confirmed the association of previously reported variants within *BCL2*, *FAS*, and *BAK1* with CLL risk. Our functional data suggest that the effect of *CDKN2A*, *BCL2*, and *FAS* loci on CLL risk is mediated by genetic variants that regulate *CDKN2A* and *ACTA2* mRNA expression levels and shape host immune responses. Finally, this study also revealed that autophagy-related variants do not seem to play a role in disease progression or patient survival. These latter findings must be confirmed in independent populations.

Acknowledgments

The authors thank all study participants who agreed to participate in the study.

This work has been funded by The Instituto de Salud Carlos III and Fondo Europeo de Desarrollo Regional (FEDER; Madrid, Spain; PI17/02256 and PI20/01845 [J.S.]; PI11/02213 and PI15/00966 [R.M.-G.]), Consejería de Transformación Económica, Industria, Conocimiento y Universidades and FEDER (PY20/01282 [J.S.]), Josep Carreras Leukaemia Research Institute (grant no. FIJC1100 [R.M.-G.]), and the voluntary economical contribution of patients. This research was also supported by the EU fund, PNRR CN3 Terapia Genica-Spoke 2 (project no. CN00000041)

and Associazione Italiana contro le Leucemie Modena Organizzazione di Volontariato (M.L.). The University of Utah was supported by funding from the National Cancer Institute (NCI) grants R01 CA134674 and P30 CA042014-29S9 (N.J.C.). Data collection in Utah was supported by the Utah Population Database (UPDB) and the Utah Cancer Registry (UCR). The UPDB is supported by the Huntsman Cancer Institute (HCI) (including Huntsman Cancer Foundation), The University of Utah, and NCI grant P30 CA2014. The UCR was additionally funded by the NCI's Surveillance, Epidemiology, and End Results Program, HHSN261201800016I, and the US Center for Disease Control and Prevention's National Program of Cancer Registries (NU58DP007131). The University of Utah thanks all study participants and the ascertainment, laboratory, biobanking, and research informatics teams at the HCI. The Interlymph Data Coordinating Center was supported by the NCI grant U01CA257679.

Authors identified as personnel of the International Agency for Research on Cancer/World Health Organization are alone responsible for the views expressed in this article, and they do not necessarily represent the decisions, policies, or views of the International Agency for Research on Cancer/World Health Organization.

Authorship

Contribution: J.S. designed and coordinated this study, performed the statistical analysis of the functional data, and obtained funding and performed data quality control together with A.J.C.-S., who performed genetic association analyses; A.J.C.-S., J.M.S.-M., P.G.-M., E.C.-C., and E.M.P., and J.S. were involved in the generation of

genetic data from the CRuCIAL cohort and drafted the manuscript; M.Á.L.-N., R.T.H., and Y.L. provided functional raw data from the Human Functional Genomic Project cohort; J.J.R.-S., F.J.R.-Z., R.C., A.P., Y.B., A.J., S.L., B.E., R.M., M.Á.L.-N., S.R.-C., C.G.-O., T.-H.C.-L., V.M., F.J., R.M.-G., M.C.-F., B.S.-M., I.G., M.G.-Á., N.J.C., T.D.-S., J.K., A.D.N., M.L., M.A., D.C., S.d.S., R.M., A.C.-G., F.C., M.I., J.M., D.C., S.I.B., and S.L.S. provided genetic data; B.S.-M. and P.L. provided autophagy flux data; and all authors contributed, edited, and approved the final version of the manuscript.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

ORCID profiles: A.J.C.-S., [0000-0003-0346-7960](https://orcid.org/0000-0003-0346-7960); J.J.R.-S., [0000-0002-4741-7925](https://orcid.org/0000-0002-4741-7925); F.J.R.-Z., [0000-0003-3438-0753](https://orcid.org/0000-0003-3438-0753); R.C., [0000-0002-8205-6192](https://orcid.org/0000-0002-8205-6192); E.C.-C., [0000-0003-2724-6759](https://orcid.org/0000-0003-2724-6759); R.T.H., [0000-0003-0576-5873](https://orcid.org/0000-0003-0576-5873); S.L., [0000-0001-8364-6357](https://orcid.org/0000-0001-8364-6357); B.E., [0000-0002-4294-8145](https://orcid.org/0000-0002-4294-8145); R. Maffei, [0000-0002-3518-2006](https://orcid.org/0000-0002-3518-2006); M.Á.L.-N., [0000-0002-3465-6062](https://orcid.org/0000-0002-3465-6062); S.R.-C., [0000-0001-9175-7102](https://orcid.org/0000-0001-9175-7102); T.-H.C.-L., [0000-0001-5969-8640](https://orcid.org/0000-0001-5969-8640); F.J., [0000-0001-7419-8492](https://orcid.org/0000-0001-7419-8492); R.M.-G., [0000-0001-9824-3657](https://orcid.org/0000-0001-9824-3657); M.C.-F., [0009-0003-8736-7146](https://orcid.org/0009-0003-8736-7146); B.S.-M., [0000-0001-6580-0971](https://orcid.org/0000-0001-6580-0971); N.J.C., [0000-0002-4788-1998](https://orcid.org/0000-0002-4788-1998); T.D.-S., [0000-0002-6127-0077](https://orcid.org/0000-0002-6127-0077); J.K., [0009-0003-5913-9576](https://orcid.org/0009-0003-5913-9576); Y.L., [0000-0003-4022-7341](https://orcid.org/0000-0003-4022-7341); R. Marasca, [0000-0002-6431-6878](https://orcid.org/0000-0002-6431-6878); P.L., [0000-0003-4130-7167](https://orcid.org/0000-0003-4130-7167); F.C., [0000-0002-4261-4583](https://orcid.org/0000-0002-4261-4583); D.C., [0000-0002-7874-3707](https://orcid.org/0000-0002-7874-3707); S.I.B., [0000-0001-5230-0652](https://orcid.org/0000-0001-5230-0652); S.L.S., [0000-0002-5173-4712](https://orcid.org/0000-0002-5173-4712); J.S., [0000-0002-9355-2423](https://orcid.org/0000-0002-9355-2423).

Correspondence: Juan Sainz, Department of Biochemistry and Molecular Biology, Faculty of Sciences, University of Granada, Avda Fuente Nueva, s/n, 18071 Granada, Spain; email: jsainz@ugr.es.

References

1. Alaggio R, Amador C, Anagnostopoulos I, et al. The 5th edition of the World Health Organization classification of haematolymphoid tumours: lymphoid neoplasms. *Leukemia*. 2022;36(7):1720-1748.
2. Hallek M. Chronic lymphocytic leukemia: 2020 update on diagnosis, risk stratification and treatment. *Am J Hematol*. 2019;94(11):1266-1287.
3. Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer statistics, 2022. *CA Cancer J Clin*. 2022;72(1):7-33.
4. Spier CM, Kjeldsberg CR, Head DR, DiFiore KC, Tudor B. Chronic lymphocytic leukemia in young adults. *Am J Clin Pathol*. 1985;84(5):675-678.
5. Ma S, Seymour JF, Brander DM, et al. Efficacy of venetoclax plus rituximab for relapsed CLL: 5-year follow-up of continuous or limited- duration therapy. *Blood*. 2021;138(10):836-846.
6. Mansouri L, Thorvaldsdottir B, Sutton LA, et al. Different prognostic impact of recurrent gene mutations in chronic lymphocytic leukemia depending on IGHV gene somatic hypermutation status: a study by ERIC in HARMONY. *Leukemia*. 2023;37(2):339-347.
7. Backman JD, Li AH, Marcketta A, et al. Exome sequencing and analysis of 454,787 UK Biobank participants. *Nature*. 2021;599(7886):628-634.
8. Jiang L, Zheng Z, Fang H, Yang J. A generalized linear mixed model association tool for biobank-scale data. *Nat Genet*. 2021;53(11):1616-1621.
9. Berndt SI, Vijai J, Benavente Y, et al. Correction: Distinct germline genetic susceptibility profiles identified for common non-Hodgkin lymphoma subtypes. *Leukemia*. 2023;37(10):2142.
10. Berndt SI, Vijai J, Benavente Y, et al. Distinct germline genetic susceptibility profiles identified for common non-Hodgkin lymphoma subtypes. *Leukemia*. 2022;36(12):2835-2844.
11. Lin WY, Fordham SE, Sunter N, et al. Genome-wide association study identifies risk loci for progressive chronic lymphocytic leukemia. *Nat Commun*. 2021;12(1):665.
12. Avsec D, Jakoš Djordjević AT, Kandušer M, Podgornik H, Škerget M, Mlinarič-Raščan I. Targeting autophagy triggers apoptosis and complements the action of venetoclax in chronic lymphocytic leukemia cells. *Cancers (Basel)*. 2021;13(18):4557.
13. Mahoney E, Maddocks K, Flynn J, et al. Identification of endoplasmic reticulum stress-inducing agents by antagonizing autophagy: a new potential strategy for identification of anti-cancer therapeutics in B-cell malignancies. *Leuk Lymphoma*. 2013;54(12):2685-2692.

14. Kopeina GS, Zhivotovsky B. The new face of autophagy in chronic lymphocytic leukemia. *Cancers (Basel)*. 2023;15(21):5279.
15. Smith LD, Minton AR, Blunt MD, et al. BCR signaling contributes to autophagy regulation in chronic lymphocytic leukemia. *Leukemia*. 2020;34(2):640-644.
16. Kong YL, Huang Y, Wu JZ, et al. Expression of autophagy related genes in chronic lymphocytic leukemia is associated with disease course. *Leuk Res*. 2018;66:8-14.
17. Arroyo DS, Rodriguez CM, Bussi C, et al. Increased expression of autophagy protein LC3 in two patients with progressing chronic lymphocytic leukemia. *Front Endocrinol (Lausanne)*. 2020;11:321.
18. Gade P, Kimball AS, DiNardo AC, et al. Death-associated protein kinase-1 expression and autophagy in chronic lymphocytic leukemia are dependent on activating transcription factor-6 and CCAAT/enhancer-binding protein-beta. *J Biol Chem*. 2016;291(42):22030-22042.
19. Bologna C, Buonincontri R, Serra S, et al. SLAMF1 regulation of chemotaxis and autophagy determines CLL patient response. *J Clin Invest*. 2016;126(1):181-194.
20. El-Khoury V, Pierson S, Szwarcbart E, et al. Disruption of autophagy by the histone deacetylase inhibitor MGCD0103 and its therapeutic implication in B-cell chronic lymphocytic leukemia. *Leukemia*. 2014;28(8):1636-1646.
21. Fonte E, Vilia MG, Reverberi D, et al. Toll-like receptor 9 stimulation can induce I kappa Bzeta expression and IgM secretion in chronic lymphocytic leukemia cells. *Haematologica*. 2017;102(11):1901-1912.
22. Su M, Mei Y, Sinha S. Role of the crosstalk between autophagy and apoptosis in cancer. *J Oncol*. 2013;2013:102735.
23. Maiuri MC, Zalckvar E, Kimchi A, Kroemer G. Self-eating and self-killing: crosstalk between autophagy and apoptosis. *Nat Rev Mol Cell Biol*. 2007;8(9):741-752.
24. Bhattacharya S, Piya S, Ma H, et al. Targeting Unc51-like autophagy activating kinase 1 (ULK1) overcomes adaptive drug resistance in acute myelogenous leukemia. *Mol Cancer Res*. 2023;21(6):548-563.
25. Salwa A, Ferraresi A, Secomandi E, et al. High BECN1 expression negatively correlates with BCL2 expression and predicts better prognosis in diffuse large B-cell lymphoma: role of autophagy. *Cells*. 2023;12(15):1924.
26. Shibusaki ST, Saitoh T, Nowag H, Münz C, Yoshimori T. Autophagy and autophagy-related proteins in the immune system. *Nat Immunol*. 2015;16(10):1014-1024.
27. Qian M, Fang X, Wang X. Autophagy and inflammation. *Clin Transl Med*. 2017;6(1):24.
28. Wei J, Long L, Yang K, et al. Autophagy enforces functional integrity of regulatory T cells by coupling environmental cues and metabolic homeostasis. *Nat Immunol*. 2016;17(3):277-285.
29. Jiang GM, Tan Y, Wang H, et al. The relationship between autophagy and the immune system and its applications for tumor immunotherapy. *Mol Cancer*. 2019;18(1):17.
30. Bakker OB, Aguirre-Gamboa R, Sanna S, et al. Integration of multi-omics data and deep phenotyping enables prediction of cytokine responses. *Nat Immunol*. 2018;19(7):776-786.
31. Li Y, Oosting M, Smeekens SP, et al. A functional genomics approach to understand variation in cytokine production in humans. *Cell*. 2016;167(4):1099-1110.e14.
32. Ter HR, Jaeger M, Smeekens SP, et al. Host and environmental factors influencing individual human cytokine responses. *Cell*. 2016;167(4):1111-1124.e13.
33. Turner JJ, Morton LM, Linet MS, et al. InterLymph hierarchical classification of lymphoid neoplasms for epidemiologic research based on the WHO classification (2008): update and future directions. *Blood*. 2010;116(20):e90-e98.
34. Pritchard JK, Stephens M, Donnelly P. Inference of population structure using multilocus genotype data. *Genetics*. 2000;155(2):945-959.
35. Kurki MI, Karjalainen J, Palta P, et al. FinnGen provides genetic insights from a well-phenotyped isolated population. *Nature*. 2023;613(7944):508-518.
36. Gagliano Taliun SA, VandeHaar P, Boughton AP, et al. Exploring and visualizing large-scale genetic associations by using PheWeb. *Nat Genet*. 2020;52(6):550-552.
37. Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics*. 2010;26(17):2190-2191.
38. Garcia-Martin P, Diez AM, Maldonado JMS, et al. Validation and functional characterization of GWAS-identified variants for chronic lymphocytic leukemia: a CRuCIAL study. *Blood Cancer J*. 2022;12(5):79.
39. Klionsky DJ, Abdel-Aziz AK, Abdelfatah S, et al. Guidelines for the use and interpretation of assays for monitoring autophagy (4th edition)(1). *Autophagy*. 2021;17(1):1-382.
40. Clavero E, Sanchez-Maldonado JM, Macaуда A, et al. Polymorphisms within autophagy-related genes as susceptibility biomarkers for multiple myeloma: a meta-analysis of three large cohorts and functional characterization. *Int J Mol Sci*. 2023;24(10):8500.
41. Aguirre-Gamboa R, Joosten I, Urbano PCM, et al. Differential effects of environmental and genetic factors on T and B cell immune traits. *Cell Rep*. 2016;17(9):2474-2487.
42. Rentsch P, Witten D, Cooper GM, Shendure J, Kircher M. CADD: predicting the deleteriousness of variants throughout the human genome. *Nucleic Acids Res*. 2019;47(D1):D886-D894.
43. Foulkes WD, Flanders TY, Pollock PM, Hayward NK. The CDKN2A (p16) gene and human cancer. *Mol Med*. 1997;3(1):5-20.

44. Landi MT, Bishop DT, MacGregor S, et al. Genome-wide association meta-analyses combining multiple risk phenotypes provide insights into the genetic architecture of cutaneous melanoma susceptibility. *Nat Genet.* 2020;52(5):494-504.
45. Rashkin SR, Graff RE, Kachuri L, et al. Pan-cancer study detects genetic risk variants and shared genetic basis in two large cohorts. *Nat Commun.* 2020;11(1):4423.
46. Vijaykrishnan J, Qian M, Studd JB, et al. Identification of four novel associations for B-cell acute lymphoblastic leukaemia risk. *Nat Commun.* 2019;10(1):5348.
47. Guney S, Jardin F, Bertrand P, et al. Several mechanisms lead to the inactivation of the CDKN2A (P16), P14ARF, or CDKN2B (P15) genes in the GCB and ABC molecular DLBCL subtypes. *Genes Chromosomes Cancer.* 2012;51(9):858-867.
48. Budina-Kolomets A, Hontz RD, Pimkina J, Murphy ME. A conserved domain in exon 2 coding for the human and murine ARF tumor suppressor protein is required for autophagy induction. *Autophagy.* 2013;9(10):1553-1565.
49. Peller S, Kaufman S. Decreased CD45RA T cells in B-cell chronic lymphatic leukemia patients: correlation with disease stage. *Blood.* 1991;78(6):1569-1573.
50. Palma M, Gentilcore G, Heimersson K, et al. T cells in chronic lymphocytic leukemia display dysregulated expression of immune checkpoints and activation markers. *Haematologica.* 2017;102(3):562-572.
51. Valvano L, Nozza F, D'Arena G, et al. Preliminary analysis of double-negative T, double-positive T, and natural killer T-like cells in B-cell chronic lymphocytic leukemia. *Cancer Med.* 2023;12(12):13241-13255.
52. Dong Y, Zheng M, Wang X, Yu C, Qin T, Shen X. High expression of CDKN2A is associated with poor prognosis in colorectal cancer and may guide PD-1-mediated immunotherapy. *BMC Cancer.* 2023;23(1):1097.
53. Zhu Z, Song H, Xu J. CDKN2A deletion in melanoma excludes T cell infiltration by repressing chemokine expression in a cell cycle-dependent manner. *Front Oncol.* 2021;11:641077.
54. Zhao Y, Zhao H, Sun Y, et al. IL-4 induces a suppressive IL-10-producing CD8+ T cell population via a Cdkn2a-dependent mechanism. *J Leukoc Biol.* 2013;94(6):1103-1112.
55. Gutiontov SI, Turchan WT, Spurr LF, et al. CDKN2A loss-of-function predicts immunotherapy resistance in non-small cell lung cancer. *Sci Rep.* 2021;11(1):20059.
56. Adib E, Nassar AH, Akl EW, et al. CDKN2A alterations and response to immunotherapy in solid tumors. *Clin Cancer Res.* 2021;27(14):4025-4035.
57. Vaux DL, Cory S, Adams JM. Bcl-2 gene promotes haemopoietic cell survival and cooperates with c-myc to immortalize pre-B cells. *Nature.* 1988;335(6189):440-442.
58. Robertson LE, Plunkett W, McConnell K, Keating MJ, McDonnell TJ. Bcl-2 expression in chronic lymphocytic leukemia and its correlation with the induction of apoptosis and clinical outcome. *Leukemia.* 1996;10(3):456-459.
59. Aviram A, Rabizadeh E, Zimra Y, et al. Expression of bcl-2 and bax in cells isolated from B-chronic lymphocytic leukemia patients at different stages of the disease. *Eur J Haematol.* 2000;64(2):80-84.
60. Dyer MJ, Zani VJ, Lu WZ, et al. BCL2 translocations in leukemias of mature B cells. *Blood.* 1994;83(12):3682-3688.
61. Berndt SI, Skibola CF, Joseph V, et al. Genome-wide association study identifies multiple risk loci for chronic lymphocytic leukemia. *Nat Genet.* 2013;45(8):868-876.
62. Hanada M, Delia D, Aiello A, Stadtmauer E, Reed JC. bcl-2 gene hypomethylation and high-level expression in B-cell chronic lymphocytic leukemia. *Blood.* 1993;82(6):1820-1828.
63. Cimmino A, Calin GA, Fabbri M, et al. miR-15 and miR-16 induce apoptosis by targeting BCL2. *Proc Natl Acad Sci U S A.* 2005;102(39):13944-13949.
64. Otake Y, Soundararajan S, Sengupta TK, et al. Overexpression of nucleolin in chronic lymphocytic leukemia cells induces stabilization of bcl2 mRNA. *Blood.* 2007;109(7):3069-3075.
65. Harigai M, Miyashita T, Hanada M, Reed JC. A cis-acting element in the BCL-2 gene controls expression through translational mechanisms. *Oncogene.* 1996;12(6):1369-1374.
66. Li C, Jiang P, Wei S, Xu X, Wang J. Regulatory T cells in tumor microenvironment: new mechanisms, potential therapeutic strategies and future prospects. *Mol Cancer.* 2020;19(1):116.
67. Schmidt A, Oberle N, Krammer PH. Molecular mechanisms of treg-mediated T cell suppression. *Front Immunol.* 2012;3:51.
68. Bauvois B, Pramil E, Jondreville L, Quiney C, Nguyen-Khac F, Susin SA. Activation of interferon signaling in chronic lymphocytic leukemia cells contributes to apoptosis resistance via a JAK-Src/STAT3/Mcl-1 signaling pathway. *Biomedicine.* 2021;9(2):188.
69. Chen Y, Shao X, Yang H, et al. Interferon gamma regulates a complex pro-survival signal network in chronic lymphocytic leukemia. *Eur J Haematol.* 2023;110(4):435-443.
70. Shao X, Meng X, Yang H, et al. IFN-gamma enhances CLL cell resistance to ABT-199 by regulating MCL-1 and BCL-2 expression via the JAK-STAT3 signaling pathway. *Leuk Lymphoma.* 2023;64(1):71-78.
71. Gregg R, Smith CM, Clark FJ, et al. The number of human peripheral blood CD4+ CD25high regulatory T cells increases with age. *Clin Exp Immunol.* 2005;140(3):540-546.

72. Baecher-Allan C, Brown JA, Freeman GJ, Hafler DA. CD4+CD25high regulatory cells in human peripheral blood. *J Immunol.* 2001;167(3):1245-1253.
73. Maffei R, Bulgarelli J, Fiorcari S, et al. The monocytic population in chronic lymphocytic leukemia shows altered composition and deregulation of genes involved in phagocytosis and inflammation. *Haematologica.* 2013;98(7):1115-1123.
74. Friedman DR, Sibley AB, Owzar K, et al. Relationship of blood monocytes with chronic lymphocytic leukemia aggressiveness and outcomes: a multi-institutional study. *Am J Hematol.* 2016;91(7):687-691.
75. Cabarcas-Petroski S, Schramm L. BDP1 alterations correlate with clinical outcomes in breast cancer. *Cancers (Basel).* 2022;14(7):1658.
76. Dicker F, Kater AP, Fukuda T, Kipps TJ. Fas-ligand (CD178) and TRAIL synergistically induce apoptosis of CD40-activated chronic lymphocytic leukemia B cells. *Blood.* 2005;105(8):3193-3198.
77. Secchiero P, Tiribelli M, Barbarotto E, et al. Aberrant expression of TRAIL in B chronic lymphocytic leukemia (B-CLL) cells. *J Cell Physiol.* 2005;205(2):246-252.
78. Olsson A, Diaz T, Aguilar-Santelises M, et al. Sensitization to TRAIL-induced apoptosis and modulation of FLICE-inhibitory protein in B chronic lymphocytic leukemia by actinomycin D. *Leukemia.* 2001;15(12):1868-1877.
79. van Attekum MHA, van Bruggen JAC, Slinger E, et al. CD40 signaling instructs chronic lymphocytic leukemia cells to attract monocytes via the CCR2 axis. *Haematologica.* 2017;102(12):2069-2076.
80. Zhang J, Jiang X, Zhang C, et al. Actin alpha 2 (ACTA2) downregulation inhibits neural stem cell migration through Rho GTPase activation. *Stem Cells Int.* 2020;2020:4764012.
81. Hoshimaru T, Nonoguchi N, Kosaka T, et al. Actin alpha 2, smooth muscle (ACTA2) is involved in the migratory potential of malignant gliomas, and its increased expression at recurrence is a significant adverse prognostic factor. *Brain Sci.* 2023;13(10):1477.
82. Kadomoto S, Izumi K, Mizokami A. The CCL20-CCR6 axis in cancer progression. *Int J Mol Sci.* 2020;21(15):5186.
83. Samaniego R, Gutiérrez-González A, Gutiérrez-Seijo A, et al. CCL20 expression by tumor-associated macrophages predicts progression of human primary cutaneous melanoma. *Cancer Immunol Res.* 2018;6(3):267-275.
84. Nüchel H, Frey UH, Bau M, et al. Association of a novel regulatory polymorphism (-938C>A) in the BCL2 gene promoter with disease progression and survival in chronic lymphocytic leukemia. *Blood.* 2007;109(1):290-297.
85. Zenz T, Benner A, Dührsen U, et al. BCL2-938C>A polymorphism and disease progression in chronic lymphocytic leukemia. *Leuk Lymphoma.* 2009;50(11):1837-1842.
86. Cabrera-Serrano AJ, Sánchez-Maldonado JM, Ter Horst R, et al. Do GWAS-identified risk variants for chronic lymphocytic leukemia influence overall patient survival and disease progression? *Int J Mol Sci.* 2023;24(9):8005.